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Amendments to the Specification:

Please replace the paragraph at page 3, lines 20-31, with the following amended

paragraph:

The thymus is arguably the major organ in the immune system because it is the

primary site of production of T lymphocytes. Its role is to attract appropriate bone

marrow-derived precursor cells from the blood, and induce their commitment to the T

cell lineage including the gene rearrangements necessary for the production of the T cell

receptor for antigen (TCR). Associated with this is a remarkable degree of cell division

to expand the number of T cells and hence increase increases the likelihood that every

foreign antigen will be recognized and eliminated. A unique feature of T cell

recognition of antigen, however, is that unlike B cells, the TCR only recognizes peptide

fragments physically associated with MHC molecules; normally this is self MHC and

this ability is selected for in the thymus. This process is called positive selection and is

an exclusive feature of cortical epithelial cells. If the TCR fails to bind to the self

MHC/peptide complexes, the T cell dies by "neglect" – it needs some degree of

signalling through the TCR for its continued maturation.

Please replace the paragraph at page 8, lines 2-8, with the following amended

paragraph:

The present inventors have demonstrated that thymic atrophy (aged induced

age-induced, or as a consequence of conditions such as chemotherapy or radiotherapy)

can be profoundly reversed by inhibition of sex steroid production, with virtually

complete restoration of thymic structure and function. The present inventors have also

found that the basis for this thymus regeneration is in part due to the initial expansion

of precursor cells which cells, which are derived both intrathymically and via the blood

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stream. This finding suggests that is possible to seed the thymus with exogenous haemopoietic stem cells (HSC) which (HSC), which have been injected into the subject.

Please replace the paragraph at page 8, lines 9-13, with the following amended paragraph:

The ability to seed the thymus with genetically modified or exogenous HSC by disrupting sex steroid signaling steroid-signaling to the thymus, means that gene therapy in the HSC may be used more efficiently to treat T cell (and myeloid cells which develop in the thymus) disorders. HSC stem cell therapy has met with little or no success to date because the thymus is dormant and incapable of taking up many if any HSC, with T cell production less than 1% of normal levels.

Please replace the paragraph at page 8, lines 15-22, with the following amended paragraph:

The present disclosure concerns methods for destroying a patient's T cells to reduce clinical disease, where the disease is related to the presence of an abnormal set of T cells. This step is followed by thymic reactivation via blockage of sex steroid mediated signaling to the thymus. The degree and kinetics of thymic regrowth can be enhanced by injection of CD34+ hematopoietic stem cells (HSC), such as autologous HSC. The patient, having been depleted of T cells, will no longer have the disease and in the presence of a rapidly reforming thymus will soon produce a new cohort of T cells in the blood and lymphoid organs thereby providing immune protection against pathogens.

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Please replace the paragraph at page 8, lines 23-27, with the following amended

paragraph:

These methods are based on disrupting sex steroid-mediated steroid-mediated signaling to the thymus in the subject. In a further embodiment, the subject is post-

pubertal. In one embodiment castration is used to disrupt the sex steroid mediated

steroid-mediated signaling. In a preferred embodiment, chemical castration is used. In

another embodiment surgical castration is used. Castration reverses the state of the

thymus to its pre-pubertal state, thereby reactivating it.

Please replace the paragraph at page 8, lines 28-34, with the following amended

paragraph:

In certain embodiments, inhibition of sex steroid production is achieved by either

castration or administration of a sex steroid analogue(s) analog(s). Non-limiting sex

steroid analogues analogs include eulexin, goserelin, leuprolide, dioxalan derivatives,

such as triptorelin, meterelin, buserelin, histrelin, nafarelin, lutrelin, leuprorelin, and

luteinizing hormone-releasing hormone analogues analogs. In some embodiments, the

sex steroid analogue analog is an analogue analog of luteinizing hormone-releasing

hormone. In certain embodiments, the luteinizing hormone-releasing hormone

analogue analog is deslorelin.

Please replace the paragraph at page 9, lines 1-4, with the following amended

paragraph:

In a particular embodiment sex steroid-mediated signaling to

the thymus is blocked by the administration of GnRH (or analogs thereof), agonists or

antagonists of LHRH, anti-estrogen antibodies, anti-androgen antibodies, passive

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(antibody) or active (antigen) anti-LHRH vaccinations, or combinations thereof

("blockers").

Please replace the paragraph at page 9, lines 13-17, with the following amended

paragraph:

In one embodiment, a patient's autoimmune disease is eliminated at least in part

by clearance of the patient's T cell population. Sex steroid mediated steroid-mediated

signaling to the thymus is disrupted. Upon regeneration of the thymus and

repopulation of the peripheral blood with new T cells, the aberrant T cells that failed to

recognize self remain eliminated from the T cell population.

Please replace the paragraph at page 9, lines 18-20, with the following amended

paragraph:

In another embodiment, a patient's allergies are eliminated by the same

disruption of sex steroid mediated steroid-mediated signaling to the thymus, followed

by regeneration of the thymus and repopulation of the peripheral blood stream with a

"clean" population of T cells.

Please replace the paragraph at page 9, lines 25-31, with the following amended

paragraph:

In cases where the subject is infected with HIV, the HSC may be genetically

modified such that they and their progeny, in particular T cells, macrophages and

dendritic cells, are resistant to infection and / or and/or destruction with the HIV virus.

The genetic modification may involve introduction into the HSC of one or more nucleic

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acid molecules which prevent viral replication, assembly and/or infection. The nucleic

acid molecule may be a gene which enclodes encodes an antiviral protein, an antisense

construct, a ribozyme, a dsRNA and a catalytic nucleic acid molecule.

Please replace the paragraph at page 10, lines 18-26, with the following amended

paragraph:

The method of the present invention is particularly relevant for treatment of

AIDS, where the treatment preferably involves reduction of viral load, reactivation of

thymic function through inhibition of sex steroids and transfer into the patients of HSC

(autologous or from a second party donor) which have been genetically modified such

that all progeny (especially T cells, DC) are resistant to further HIV infection. This

means that not only will the patient be depleted of HIV virus and no longer susceptible

to general infections because the T cells have returned to normal levels, but the new T

cells being resistant to HIV will be able to remove any remnant viral infected cells. In

principle a similar strategy could be applied to gene therapy in HSC for any T cell

defect or any viral infection which targets T cells.

Please replace the paragraph at page 25, lines 5-7, with the following amended

paragraph:

Figures 47A-47C are bar graphs showing that there was a significant increase in

all thymocyte subclasses (Fig. 47A) in castrated NOD mice. There was no change in B

cells compared to sham-castrated NOD mice (Fig. 47C) nor in the total T or

B cells in the spleen (Fig. 47B).

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Please replace the paragraph at page 27, lines 18-23, with the following amended paragraph:

The recipient's thymus may be reactivated by disruption of sex steroid mediated signalling steroid-mediated signaling to the thymus. This disruption reverses the hormonal status of the recipient. In certain embodiments, the recipient is post-pubertal. According to the methods of the invention, the hormonal status of the recipient is reversed such that the hormones of the recipient approach pre-pubertal levels. By lowering the level of sex steroid hormones in the recipient, the signalling of these hormones to the thymus is lowered, thereby allowing the thymus to be reactivated.

Please replace the paragraph at page 27, lines 24-30, with the following amended paragraph:

A non-limiting method for creating disruption of sex steroid mediated signalling steroid-mediated signaling to the thymus is through castration. Methods for castration include, but are not limited to, chemical castration and surgical castration. During or after the castration step, hematopoietic stem or progenitor cells, or epithelial stem cells, from the donor are transplanted into the recipient. These cells are accepted by the thymus as belonging to the recipient and become part of the production of new T cells and DC by the thymus. The resulting population of T cells recognize both the recipient and donor as self, thereby creating tolerance for a graft from the donor.

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Please replace the paragraph at page 29, lines 17-30, with the following amended paragraph:

Administration may be by any method which delivers the sex steroid ablating steroid-ablating agent into the body. Thus, the sex steroid ablating sex steroid-ablating agent maybe be may be administered, in accordance with the invention, by any route including, without limitation, intravenous, subdermal, subcutaneous, intramuscular, topical, and oral routes of administration. One non-limiting example of administration of a sex steroid-ablating steroid-ablating agent is a subcutaneous/intradermal injection of a "slow-release" depot of GnRH agonist (e.g., one, three, or four month Lupron® injections) or a subcutaneous/intradermal injection of a "slow-release" GnRHcontaining implant (e.g., one or three month Zoladex®, e.g., 3.6 mg or 10.8 mg implant). These could also be given intramuscular intramuscularly (i.m.), intravenously (i.v.) or orally, depending on the appropriate formulation. Another example is by subcutaneous injection of a "depot" or "impregnated implant" containing, for example, about 30 mg of Lupron® (e.g., Lupron Depot®, (leuprolide acetate for depot suspension) TAP Pharmaceuticals Products, Inc., Lake Forest, IL) (e.g., Lupron Depot®, (leuprolide acetate for depot suspension) TAP Pharmaceutical Products, Inc., Lake Forest, IL). A 30 mg Lupron® injection is sufficient for four months of sex steroid ablation to allow the thymus to rejuvenate and export new naïve T cells into the blood stream.

Please replace the paragraph at page 30, lines 1-25, with the following amended paragraph:

In some embodiments, sex steroid ablation or inhibition of sex steroid signaling steroid-signaling is accomplished by administering an anti-androgen such as an

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androgen blocker (e.g., bicalutamide, trade names Cosudex® or Casodex®, AstraZeneca, Aukland Auckland, NZ), either alone or in combination with an LHRH analog or any other method of castration. Sex steroid ablation or interruption of sex steroid-signalling steroid-signaling may also be accomplished by administering cyproterone acetate (trade name, Androcor®, Shering Schering AG, Germany; e.g., 10-1000 mg, 100 mg bd or tds, or 300 mg IM weekly, a 17-hydroxyprogesterone acetate, which acts as a progestin, either alone or in combination with an LHRH analog or any other method of castration. Alternatively, other anti-androgens may be used (e.g., antifungal agents of the imidazole class, such as liarozole(Liazol® e.g., 150 mg/day, an aromatase inhibitor) liarozole (Liazol®, e.g., 150 mg/day, an aromatase inhibitor) and ketoconazole, bicalutamide (trade name Cosudex® or Casodex®, 5-500 mg, e.g., 50 mg po QID), flutamide (trade names Euflex® and Eulexin®, Shering Schering Plough Corp, N.J.; 50-500 mg e.g., 250 or 750 po QID), megestrol acetate (Megace®) e.g., 480-840 mg/day or nilutamide (trade names Anandron®, and Nilandron®, Roussel, France e.g., orally, 150-300 mg/day)). Antiandrogens are often important in therapy, since they are commonly utilized to address flare by GnRH analogs. Some antiandrogens act by inhibiting androgen receptor translocation, which interrupts negative feedback resulting in increased testosterone levels and minimal loss of libido/potency. Another class of anti-androgens useful in the present invention are the selective androgen receptor modulators (SARMS) (e.g., quinoline derivatives, bicalutamide (trade name Cosudex® or Casodex®, ICI Pharmaceuticals, England e.g., orally, 50 mg/day), and flutamide (trade name Eulexin®, e.g., orally, 250 mg/day)). Other well known antiandrogens include 5 alpha reductase inhibitors (e.g., dutasteride, (e.g., 0.5 mg/day) dutasteride, (e.g., 0.5 mg/day) which inhibits both 5 alpha reductase isoenzymes and results in greater and more rapid DHT suppression; finasteride (trade name Proscar®; 0.5-500mg e.g., 0.5-500 mg, e.g., 5 mg po daily), which inhibits 5alpha 5 alpha reductase

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2 and consequent DHT production, but has little or no effect on testosterone or LH levels).

Please replace the paragraph bridging pages 30 and 31, with the following amended paragraph:

In other embodiments, sex steroid ablation or inhibition of sex steroid signaling steroid-signaling is accomplished by administering anti-estrogens either alone or in combination with an LHRH analog or any other method of castration. Some antiestrogens (e.g., anastrozole (trade name Arimidex®), and fulvestrant (trade name Faslodex®) act by binding the estrogen receptor (ER) with high affinity similar to estradiol and consequently inhibiting estrogen from binding. Faslodex® binding also triggers conformational change to the receptor and down-regulation of estrogen receptors, without significant change in FSH or LH levels. Other non-limiting examples of anti-estrogens are tamoxifen (trade name Nolvadex®); Clomiphene (trade-name Clomid®)e.g.,50-250mg/day Clomiphene (trade name Clomid®) e.g., 50-250 mg/day, a non-steroidal ER ligand with mixed agonist/antagonist properties, which stimulates release of gonadotrophins; Fulvestrant (trade name Faslodex®; 10-1000 mg, e.g., 250mg 250 mg IM monthly); diethylstilbestrol ((DES), trade name Stilphostrol®) e.g., 1-3mg/day e.g., 1-3 mg/day, which shows estrogenic activity similar to, but greater than, that of estrone, and is therefore considered an estrogen agonist, but binds both androgen and estrogen receptors to induce feedback inhibition on FSH and LH production by the pituitary, diethylstilbestrol diphosphate e.g., 50 to 200 mg/day e.g., 50 to 200 mg/day; as well as danazol, , droloxifene <u>danazol, droloxifene</u>, and iodoxyfene, which each act as antagonists. Another class of anti-estrogens which may be used either alone or in combination with other methods of castration, are the selective

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estrogen receptor modulators (SERMS) (*e.g.*, toremifene (trade name Fareston®, 5-1000 mg, *e.g.*, 60mg 60 mg po QID), raloxofene (trade name Evista®), and tamoxifen (trade name Nolvadex®, 1-1000mg 1-1000 mg, *e.g.*, 20mg 20 mg po bd), which behaves as an agonist at estrogen receptors in bone and the cardiovascular system, and as an antagonist at estrogen receptors in the mammary gland). Estrogen receptor downregulators (ERDs) (*e.g.*, tamoxifen (trade name, Nolvadex®)) may also be used in the present invention.

Please replace the paragraph bridging pages 31 and 32, with the following amended paragraph:

Other non-limiting examples of methods of inhibiting sex steroid signalling steroid-signaling which may be used either alone or in combination with other methods of castration, include aromatase inhibitors and other adrenal gland blockers (e.g., Aminoglutethimide, formestane, vorazole, exemestane, anastrozole (trade name Arimidex®, 0.1-100 mg, e.g., 1 mg po QID), which lowers estradiol and increases LH and testosterone), letrozole (trade name Femara®, 0.2-500 mg, e.g., 2.5 mg po QID), and exemestane (trade name Aromasin®)1-2000mg, e.g., 25mg/day) (trade name Aromasin®) 1-2000 mg, e.g., 25 mg/day); aldosterone antagonists (e.g., spironolactone (trade name, Aldactone®) e.g., 100 to 400mg/day 100 to 400 mg/day), which blocks the androgen cytochrome P-450 receptor;) and eplerenone, a selective aldosterone-receptor antagonist) antiprogestogens (e.g., medroxypregesterone acetate, e.g.,5mg/day e.g., 5 mg/day, which inhibits testosterone syntheses and LH synthesis); and progestins and anti-progestins such as the selective progesterone response modulators (SPRM) (e.g., megestrol acetate e.g.,160mg/day e.g., 160 mg/day, mifepristone (RU 486, Mifeprex®, e.g.,200mg/day e.g., 200 mg/day); and other

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compounds with estrogen/antiestrogenic activity, (e.g., phytoestrogens, flavones, isoflavones and coumestan derivatives, lignans, and industrial compounds with phenolic ring (e.g., DDT)). Also, anti-GnRH vaccines (see, e.g., Hsu et al., (2000) Cancer Res. 60:3701; Talwar, (1999) Immunol. Rev. 171:173-92), or any other pharmaceutical which mimics the effects produced by the aforementioned drugs, may also be used. In addition, steroid receptor based modulators, which may be targeted to be thymic specific, may also be developed and used. Many of these mechanisms of inhibiting sex steroid signaling steroid-signaling are well known. Each drugs drug may also be used in modified form, such as acetates, citrates and other salts thereof, which are well known to those in the art.

Please replace the paragraph at page 32, lines 7-12, with the following amended paragraph:

Because of the complex and interwoven feedback mechanisms of the hormonal system, administration of sex steroids may result in inhibition of sex steroid signalling steroid-signaling. For example, estradiol decreases gonadotropin production and sensitivity to GnRH action. However, higher levels of estradiol result in gonadotropin surge. Likewise, progesterone influences frequency and amount of LH release. In men, testosterone inhibits gonadotropin production. Estrogen administered to men decreases LH and testosterone, and anti-estrogen increases LH.

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Please replace the paragraph at page 32, lines 17-21, with the following amended paragraph:

In some embodiments, the sex steroid mediated steroid-mediated signaling to the thymus is disrupted by administration of gonadotrophin-releasing hormone (GnRH) or an analog thereof. GnRH is a hypothalamic decapeptide that stimulates the secretion of the pituitary gonadotropins, leutinizing hormone (LH) and follicle-stimulating hormone (FSH). Thus, GnRH, e.g., in the form of Synarel or Lupron, will suppress the pituitary gland and stop the production of FSH and LH.

Please replace the paragraph bridging pages 32-34, with the following amended paragraph:

In some embodiments, the sex steroid mediated steroid-mediated signaling to the thymus is disrupted by administration of a sex steroid analog, such as an analog of leutinizing hormone-releasing hormone (LHRH). Sex steroid analogs and their use in therapies and chemical castration are well known. Sex steroid analogs are commercially known and their use in therapies and chemical castration are well known. Such analogs include, but are not limited to, the following agonists of the LHRH receptor (LHRH-R): buserelin (e.g., buserelin acetate, trade names Suprefact® (e.g., 0.5-02 mg s.c./day), Suprefact Depot®, and Suprefact® Nasal Spray (e.g., 2 µg per nostril, every 8 hrs.), Hoechst, also described in U.S. Patent Nos. 4,003,884, 4,118,483, and 4,275,001); Cystorelin® (e.g., gonadorelin diacetate tetrahydrate, Hoechst); deslorelin (e.g., desorelin deslorelin acetate, Deslorell®, Balance Pharmaceuticals); gonadorelin (e.g., gonadorelin hydrocholoride, trade name Factrel® (100 µg i.v. or s.c.), Ayerst Laboratories); goserelin (goserelin acetate, trade name Zoladex®, AstraZeneca, Aukland

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Auckland, NZ, also described in U.S. Patent Nos. 4,100,274 and 4,128,638; GB 9112859 and GB 9112825); histrelin (e.g., histerelin acetate histrelin acetate, Supprelin®, (s.c., 10 μg/kg.day s.c., 10 μg/kg/day, Ortho, also described in EP 217659); leuprolide (leuprolide acetate, trade name Lupron® or Lupron Depot®; Abbott/TAP, Lake Forest, IL, also described in U.S. Patent Nos. 4,490,291 3,972,859, 4,490,291, 3,972,859, 4,008,209, 4,992,421, and 4,005,063; DE 2509783); leuprorelin (e.g., leuproelin leuprorelin acetate, trade name Prostap SR® (e.g., single 3.75 mg dose s.c. or i.m./month), Prostap3® (e.g., single 11.25 mg dose s.c. every 3 months), Wyeth, USA, also described in Plosker et al., (1994) Drugs 48:930); lutrelin (Wyeth, USA, also described in U.S. Patent No. 4,089,946); Meterelin® (e.g., Avorelina (e.g., 10-15 mg slow-release formulation), also described in EP 23904 and WO 91/18016); nafarelin (e.g., trade name Synarel® (i.n. 200-1800 μg/day), Syntex, also described in U.S. Patent No. 4,234,571; W0 93/15722 <u>WO</u> 93/15722; and EP 52510 EP0052510); and triptorelin (e.g., triptorelin pamoate; trade names Trelstar LA® (11.25 mg over 3 months), Trelstar LA Debioclip® (pre-filled, single dose delivery), LA Trelstar Depot® (3.75 mg over one month), and Decapeptyl®, Debiopharm S.A., Switserland, also described in U.S. Patent Nos. 4,010,125, 4,018,726, 4,024,121, and 5,258,492; EP 364819). LHRH analogs also include, but are not limited to, the following antagonists of the LHRH-R: abarelix (trade name Plenaxis™ (e.g., 100 mg i.m. on days 1, 15 and 29, then every 4 weeks thereafter), Praecis Pharmaceuticals, Inc., Cambridge, MA) and cetrorelix (e.g., cetrorelix acetate, trade name Cetrotide™ (e.g., 0.25 or 3 mg s.c.), Zentaris, Frankfurt, Germany). Additional sex steroid analogs include Eulexin® (e.g., flutamide (e.g., 2 capsules 2x/day, total 750 mg/day), Schering-Plough Corp., also described in FR 7923545, WO 86/01105 and PT 100899), and dioxane derivatives (e.g., those described in EP 413209), and other LHRH analogues analogs such as are described in EP 181236, U.S. Patent Nos. 4,608,251, 4,656,247, 4,642,332, 4,010,149, 3,992,365, and 4,010,149. Combinations of agonists,

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combinations of antagonists, and combinations of agonists and antagonists are also included. One non-limiting analog of the invention is deslorelin (described in U.S. Patent No. 4,218,439). For a more extensive list, list of analogs, see Vickery et al. (1984) LHRH and Its Analogs: Contraceptive & Therapeutic Applications (Vickery et al., eds.) MTP Press Ltd., Lancaster, PA. Each analog may also be used in modified form, such as acetates, citrates and other salts thereof, which are well known to those in the art.

Please replace the paragraph at page 37, lines 9-23, with the following amended paragraph:

The intracellular receptors are members of the nuclear receptor superfamily. They are located in the cytoplasm of the cell and are transported to the nucleus after binding with the sex steroid hormone where they alter the transcription of specific genes. Receptors for the sex steroid hormones exist in several forms. Well known in the literature are two forms of the progesterone receptor, PRA and PRB, and three forms of the estrogen receptor, ERα, ERβ1 and ERβ2. Transcription of genes in response to the binding of the sex steroid hormone receptor to the steroid response element in the promoter region of the gene can be modified in a number of ways. Co-activators and co-repressors exist within the nucleus of the target cell that can modify binding of the steroid-receptor complex to the DNA and thereby effect transcription. The identity of many of these co-activators and co-repressors are known and methods of modifying their actions on steroid receptors are the topic of current research. Examples of the transcription factors involved in sex steroid hormone action are NF-1, SP1, Oct-1 and Oct-1 and TFIID. These co-regulators are required for the full action of the steroids. Methods of modifying the actions of these nuclear regulators could involve the balance

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between activator and repressor by the use of antagonists or through control of expression of the genes encoding the regulators.

Please replace the paragraph at page 53, lines 10-30, with the following amended paragraph:

Thus, in accordance with the invention, the basic principle is to stop ongoing autoimmune disease or prevent one developing in highly predictive cases (e.g., in familial distribution) with T cell and B cell, as appropriate, depletion followed by rebuilding a new tolerant immune system. First, the autoimmune disease is diagnosed, and a determination is made as to whether or not there is a familial (genetic) predisposition. Next, a determination is made as to whether or not there had been a recent prolonged infection in the patient which may have lead to the autoimmune disease through antigen mimicry or inadvertent clonal activation. In practice it may be impossible to determine the cause of the disease. Next, T cell depletion is performed and, as appropriate, B cell depletion is performed, combined with chemotherapy, radiation therapy or anti-B cell reagents (e.g., CD19, CD20, and CD21) or antibodies to specific Ig subclasses (anti IgE). The thymus and bone marrow function is then reactivated by administering GnRH to the patient. Simultaneous with this reactivation of the thymus and bone marrow is the injection of HSC which have been in vitro transfected with a gene encoding the autoantigen to enter the rejuvenating thymus and convert to DC for presentation of the autoantigen to developing T cells thereby inducing tolerance. The transfected HSC will also produce the antigen in the bone marrow, and present the antigen to developing immature B cells, thereby causing their deletion, similar to that occurring to T cells in the thymus. Use of the immunosuppressive regimes (anti-T, -B therapy) would overcome any untoward

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activation of pre-existing potentially autoreactive T and B cells. Moreover, in the case of no-obvious genetic predisposition, the thymic and marrow reactivation with GnRH

may be combined with G-CSF injection to increase blood levels of autologous HSC to

enhance the thymic regrowth.

Please replace the paragraph at page 54, lines 7-12, with the following amended

paragraph:

Animals. CBA/CAH and C57Bl6/J male mice were obtained from Central

Animal Services, Monash University and were housed under conventional conditions.

C57Bl6/J Ly5.1+ were obtained from the Central Animal Services Monash University

Central Animal Services, Monash University, the Walterand Walter and Eliza Hall

Institute for Medical Research (Parkville, Victoria) and the A.R.C. (Perth Western

Australia) and were housed under conventional conditions. Ages ranged from 4-6

weeks to 26 months of age and are indicated where relevant.

Please replace the paragraph at page 59, lines 5-12, with the following amended

paragraph:

The DN subpopulation, in addition to the thymocyte precursors, contains

(αβTCR-+CD4-CD8- <u>αβTCR+CD4-CD8-</u> thymocytes, which are thought to have

downregulated both co-receptors at the transition to SP cells (Godfrey & Zlotnik, 1993).

By gating on these mature cells, it was possible to analyze the true TN compartment

(CD3 CD4 CD8-) and their subpopulations expressing CD44 and CD25. Figures 5H, 5I,

5J, and 5K illustrate the extent of proliferation within each subset of TN cells in young,

old and castrated mice. This showed a significant (p<0.001) decrease in proliferation of

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the TN1 subset (CD44+CD25- CD3-CD4-CD8-), from ~10%% 10% in the normal young to around 2% at 18 months of age (Fig. 5H) which was restored by 1 week post-castration.

Please replace the paragraph at page 60, lines 20-25, with the following amended paragraph:

The thymic extracellular matrix, containing important structural and cellular adhesion molecules such as collagen, laminin and fibrinogen, was detected by the mAb MTS 16. Scattered throughout the normal young thymus, the nature of MTS 16 expression became more widespread and interconnected in the aged thymus. Expression of MTS 16 was increased further at 2 weeks post-castration while <u>at 4 weeks post-castration</u>, this expression was representative of the situation in the 2 month thymus (data not shown).

Please replace the paragraph at page 65, lines 22-29, with the following amended paragraph:

The above findings indicate a defect in the thymic epithelium rendering it rendering it incapable of providing the developing thymocytes with the necessary stimulus for, development for development. However, the symbiotic nature of the thymic, epithelium thymic epithelium and thymocytes makes it difficult to ascertain the exact pathway of destruction by the sex steroid influences. The medullary epithelium requires cortical T cells for its proper development and maintenance. Thus, if this population is diminished, the diminished, the medullary thymocytes may not receive adequate signals for development. This particularly seems to affect the CD8+

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population. IRF-/- mice show a decreased number of CD8+ T cells. It would therefore, be interesting to determine the proliferative capacity of these cells.

Please replace the paragraph at page 71, lines 16-26, with the following amended paragraph:

In both irradiation and cyclophosphamide models of immunodepletion thymocyte numbers peaked at every two weeks and decreased four weeks after treatment. Almost immediately after irradiation or chemotherapy, thymus weight and cellularity decreased dramatically and approximately 5 days later the first phase of thymic regeneration begun. The first wave of reconstitution (days 5-14) was brought about by the proliferation of radioresistant thymocytes (predominantly double negatives) which gave rise to all thymocyte subsets (Penit and Ezine 1989). The second decrease, observed between days 16 and 22 was due to the limited proliferative ability of the radioresistant cells coupled with a decreased production of thymic precursors by the bone marrow (also effected by irradiation). The second regenerative phase was due to the replenishment of the thymus with bone marrow derived precursors (Huiskamp *et al.*, 1983).

Please replace the paragraph at page 75, lines 1-14, with the following amended paragraph:

In noncastrated mice, there was a profound decrease in thymocyte number over the 4 week time period, with little or no evidence of regeneration (Fig. 21A). In the castrated group, however, by two weeks there was already extensive thymopoiesis which by four weeks had returned to control levels, being 10 fold higher than in noncastrated mice. Flow cytometeric analysis of the thymii with respect to CD45.2

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(donor-derived antigen) demonstrated that no donor derived donor-derived cells were detectable in the noncastrated group at 4 weeks, but remarkably, virtually all the thymocytes in the castrated mice were donor-derived at this time point (Fig. 21B). Given this extensive enhancement of thymopoiesis from donor-derived haemopoietic precursors, it was important to determine whether T cell differentiation had proceeded normally. CD4, CD8 and TCR defined subsets were analysed analyzed by flow cytometry. There were no proportional differences in thymocytes subset proportions 2 weeks after reconstitution (Fig. 22). This observation was not possible at 4 weeks, because the noncastrated mice were not reconstituted with donor-derived cells. However, at this time point the thymocyte proportions in castrated mice appear normal.

Please replace the paragraph at page 80, lines 11-18, with the following amended paragraph:

The patient was given sex steroid ablation therapy in the form of delivery of an LHRH agonist. This was given in the form of either Leucrin (depot injection; 22.5mg 22.5 mg) or Zoladex (implant; 10.8 mg), either one as a single dose effective for 3 months. This was effective in reducing sex steroid levels sufficiently to reactivate the thymus. In other words, the serum levels of sex steroids were undetectable (castrate; <0.5 ng/ml <0.5 ng/ml blood). In some cases it is also necessary to deliver a suppresser of adrenal gland production of sex steroids. Cosudex (5mg/day 5 mg/day) may be delivered as one tablet per day for the duration of the sex steroid ablation therapy. Adrenal gland production of sex steroids makes up around 10-15% of a human's steroids.

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Please replace the paragraph at page 80, lines 19-24, with the following amended paragraph:

Reduction of sex steroids in the blood to minimal values took about 1-3 weeks; concordant with this was the reactivation of the thymus. In some cases it is necessary to extend the treatment to a second 3 month injection/implant. The thymic expansion may be increased by simultaneous enhancement of blood HSC either as an allogeneic donor (in the case of grafts of foreign tissue) or autologous HSC (by injecting the host with G-CSF to mobilize these HSC from the bone marrow to the thymus).

Please replace the paragraph bridging pages 81 and 82, with the following amended paragraph:

Where practical, the level of hematopoietic stem cells (HSC) in the donor blood is enhanced by injecting into the donor granulocyte-colony stimulating factor (G-CSF) at 10μg/kg 10 μg/kg for 2-5 days prior to cell collection (e.g., one or two injections of 10μg/kg 10 μg/kg per day for each of 2-5 days). CD34* donor cells are purified from the donor blood or bone marrow, such as by using a flow cytometer or immunomagnetic beading. Antibodies that specifically bind to human CD34 are commercially available (from, e.g., Research Diagnostics Inc., Flanders, NJ). Donor-derived HSC are identified by flow cytometry as being CD34*. These CD34+ HSC may also be expanded by in vitro culture using feeder cells (e.g., fibroblasts), growth factors such as stem cell factor (SCF), and LIF to prevent differentiation into specific cell types. At approximately 3-4 weeks post LHRH agonist delivery (i.e., just before or at the time the thymus begins to regenerate) the patient is injected with the donor HSC, optimally at a dose of about 2-4 x 106 cells/kg. G-CSF may also be injected into the recipient to assist in expansion of the donor HSC. If this timing schedule is not possible because of the critical nature of

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clinical condition, the HSC could be administered at the same time as the GnRH. It may be necessary to give a second dose of HSC 2-3 weeks later to assist in the thymic regrowth and the development of donor DC (particularly in the thymus). Once the HSC have engraftment engrafted (i.e., have incorporated into the bone marrow and thymus), the effects should be permanent since the HSC are self-renewing.

Please replace the paragraph at page 87, lines 5-14, with the following amended paragraph:

To stop the ongoing autoimmune disease, the patient will to undergo T cell depletion. She will also undergo thymic regeneration to replace these T cells and hence overcome the immunodeficiency state. To do this, she will receive 4 one monthly injections of Lupron (7.5mg 7.5 mg) to deplete the sex steroids (by 3 weeks) thereby allowing reactivation of her thymus. This will also allow uptake of the HSC and to establish central tolerance to the autoantigen in question. It is not clear why autoimmune disease starts but cross-reaction to a microorganism is a likely possibility; depleting all T cells will thus remove these cross-reactive cells. If the disease was initiated by such cross-reaction if may not be necessary to transfect the HSC with the nominal autoantigen. Simply depleting T cells followed by thymic reactivation by disrupting sex steroid signaling to the thymus may be sufficient.

Please replace the paragraph at page 87, lines 15-25, with the following amended paragraph:

One standard procedure for removing T cells is as follows. The human patient receives anti-T cell antibodies in the form of a daily injection of 15 mg/kg of Atgam (xeno anti-T cell globulin, Pharmacia Upjohn) for a period of 10 days in

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combination with an inhibitor of T cell activation, cyclosporin A, 3mg/kg 3 mg/kg, as a continuous infusion for 3-4 weeks followed by daily tablets at 9mg/kg 9 mg/kg as needed. This treatment does not affect early T cell development in the patient's thymus, as the amount of antibody necessary to have such an effect cannot be delivered due to the size and configuration of the human thymus. The treatment is maintained for approximately 4-6 weeks to allow the loss of sex steroids followed by the reconstitution of the thymus. The prevention of T cell reactivity may also be combined with inhibitors of second level signals such as interleukins, accessory molecules (blocking, e.g., CD28), signal transduction molecules or cell adhesion molecules to enhance the T cell ablation.